

## AMENDMENTS TO THE SPECIFICATION

Please amend the specification, paragraph beginning on page 11, line 22, as follows:

Nucleic acid hybridization will be affected by such conditions as salt concentration, temperature, or organic solvents, in addition to the base composition, length of the complementary strands, and the number of nucleotide base mismatches between the hybridizing nucleic acids, as will be readily appreciated by those skilled in the art. Stringent temperature conditions will generally include temperatures in excess of ~~[[30EC]]~~ 30°C, typically in excess of ~~[[37EC]]~~ 37°C, and preferably in excess of ~~[[45EC]]~~ 45°C. Stringent salt conditions will ordinarily be less than 1000 mM, typically less than 500 mM, and preferably less than 200 mM. However, the combination of parameters is much more important than the measure of any single parameter. The stringency conditions are dependent on the length of the nucleic acid and the base composition of the nucleic acid, and can be determined by techniques well known in the art. See, e.g., Asubel, 1992; Wetmur and Davidson, 1968.

Please amend the specification, paragraph beginning on page 12, line 1, as follows:

Thus, as herein used, the term "stringent conditions" means hybridization will occur only if there is at least 95% and preferably at least 97% identity between the sequences. Such hybridization techniques are well known to those of skill in the art. Stringent hybridization conditions are as defined above or, alternatively, conditions under overnight incubation at ~~[[42EC]]~~ 42°C in a solution comprising: 50% formamide, 5x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH7.6), 5x Denhardt's solution, 10% dextran sulfate, and 20 µg/ml denatured, sheared salmon sperm DNA, followed by washing the filters in 0.1x SSC at about ~~[[65EC]]~~ 65°C.

Please amend the specification, paragraph beginning on page 24, line 23, as follows:

*In Situ Hybridizations:* *In situ* hybridization was performed as described previously (Oxtoby and Jowett, 1993). The GFP probe was made by antisense transcription of TOPdGFP, using a T7 promoter present in the construct. ~~[[Probe]]~~ The probe for *hdl* was made from a full-length clone isolated in our laboratory, subcloned into pCS2+. Probes for *pax2.1* (Krauss et al., 1991), *thx6* (Hug et al., 1997), and *lef1* (Dorsky et al., 1999) were made as described previously. Images were taken with a Kodak DC290 digital camera and processed with Adobe Photoshop 5.0.